

## NEW NATURAL ROTENOID AND PTEROCARPANOID ANALOGUES FROM *NEORAUTANENIA AMBOENSIS*

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**Key Word Index**—*Neorautanenia amboensis*; Leguminosae; rotenoids; pterocarpanoids.

**Abstract**—Four new natural rotenoid and pterocarpanoid analogues, neobanone, rotenonone, 12a-hydroxyisomillettone and neobanol were isolated from the benzene extract of the root of *Neorautanenia amboensis*. The structures were determined by investigation of IR, UV, MS,  $^1\text{H-NMR}$ , CD and oxidative conversions.

### INTRODUCTION

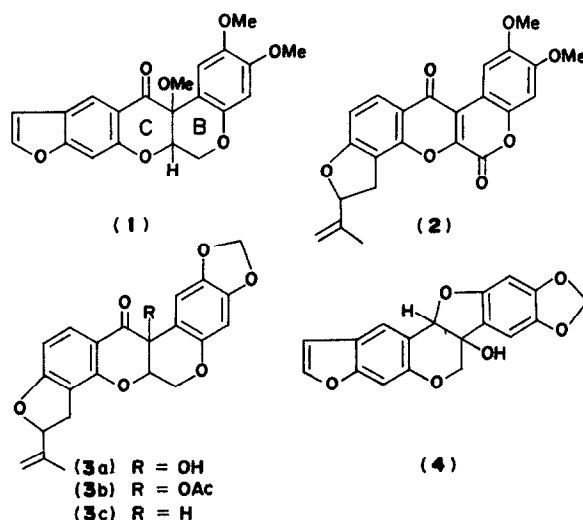
The isolation of rotenone, dolineone and their 12a-hydroxy- and 12a-O-methyl derivatives from the light petroleum extract of the root of *Neorautanenia amboensis* Schinz was described earlier [1]. Benzene extracts gave in addition three new rotenoids (1), (2) and (3a) as well as a new 6a-hydroxypterocarpan (4). Although only trace amounts (ca 1.0% of the crude extract) of these are present, spectrometry and simple oxidative conversions of biogenetic significance assist structure elucidations. Preliminary tests show (3) to be toxic to insects.

### RESULTS AND DISCUSSION

Neobanone (1) has the composition  $\text{C}_{21}\text{H}_{18}\text{O}_7$ ,  $M^+$  382,  $[\alpha]_D^{25} + 61^\circ$ . The pale yellow oil gave a purple colour with the Durham test [2], which indicates a possible rotenoid structure. The nature of the structure of neobanone was indicated by its NMR spectrum which also confirmed the presence of an ABC system (arising from the  $6_{ax,eq}$ , 6a protons— $\tau$  5.37, m,  $H_{6_{ax,eq}}$ ; 5.03, m,  $H_{6a}$ ) and provided evidence to the 12a-substituted nature of (1). The position of the 12a-O-methyl group ( $\tau$  6.44, s, 3H) is in agreement with that of similar compounds [1], while the MS was in full agreement with the proposed structure. The *cis* relationship at the B/C ring junction and absolute configuration (6aS, 12aS) was established from NMR [3] and CD data.

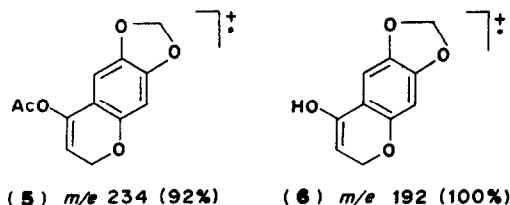
Rotenonone (2) crystallized as bright yellow needles from dichloroethane, mp 298–305° (decomp.),  $M^+$  406,  $\nu_{\text{max}}^{\text{KBr}}$  1750 (lactone C=O), 1630 (unsat. C=O)  $\text{cm}^{-1}$ . Oxidation of dehydrorotenone with *n*-amyl nitrite in acetic acid gave a product identical to the natural compound. The NMR spectrum was also in full agreement with that of the  $\text{MnO}_2$  oxidation product, mp 298–300. 5° (decomp.) of dehydrorotenone obtained by Carlson *et al.* [4].

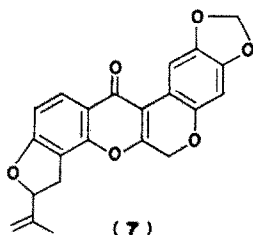
12a-Hydroxyisomillettone (3a), isolated as the acetate (3b) as light brown oil gave  $[\alpha]_D^{25} - 192$ ,  $\nu_{\text{max}}^{\text{CHCl}_3}$  1745 (ace-



tyl (C=O), 1690 (C=O)  $\text{cm}^{-1}$ . The compound analyzed for the empirical formula  $\text{C}_{22}\text{H}_{18}\text{O}_7$  in agreement with the MS. Fragments (5) and (6) suggest 12a-substitution [5] for (3b). This was confirmed by the NMR spectrum and the acid acatalyzed dehydration of (3b) to (7), yellow oil,  $\nu_{\text{max}}^{\text{CHCl}_3}$  1675 (unsat. C=O)  $\text{cm}^{-1}$ ,  $\tau$  5.43 ( $H_6$ , s, 2H). Isomillettone (3c) was previously isolated as a mixture from *Piscidia erythrina* L. [6].

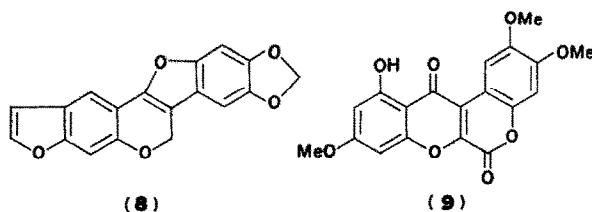
Neobanol (4),  $\text{C}_{18}\text{H}_{12}\text{O}_6$ ,  $M^+$  324,  $[\alpha]_D^{25} - 246^\circ$  was obtained as colourless needles, mp 244–246°C from ace-





tone. The IR spectrum showed no bands in the carbonyl region. The NMR spectrum indicated the presence of a furan ring (pair of doublets,  $\tau$  2.27 and 3.24, J2.2 Hz), methylenedioxy group at  $\tau$  4.09, s, and a hydroxy group ( $\tau$  5.00, br. s, exchangeable with D<sub>2</sub>O). The tertiary nature of the hydroxy group was indicated by the conversion of (4) to neoduleen (8) [7]. The downfield shift of the  $\delta_{\text{ax,eq}}$  protons from  $\tau$  5.46 (s, 2H) in (4) to 4.46 (s, 2H) in (8) is in agreement with the conversion of 6a-hydroxypterocarpan to pterocarpenes [8]. The *cis* relationship and absolute configuration (6aR, 11aR) were determined by NMR and optical rotation measurements in comparison with known structures.

The isolation of stemonone (9) [9] and rotenonone (2) as natural products is of biogenetic interest because the association of (2) with known rotenoids in the same plant [1] may be indicative of the biogenetic sequence (rotenone  $\rightarrow$  12a-hydroxyrotenone  $\rightarrow$  dehydrorotenone  $\rightarrow$  rotenonone) in their formation.



#### EXPERIMENTAL

Mp's are uncorrected. IR spectra were recorded in KBr discs or CHCl<sub>3</sub> soln. NMR were determined using a Varian T-60 spectrometer and MS were recorded on a Varian CH-5 instrument. CD spectra were measured in MeOH on a Jasco J-20 spectropolarimeter. Chemical shifts are expressed in ppm downfield from TMS. Preparative TLC was carried out on Si gel 60 PF<sub>254</sub> plates (1 mm thick).

**Isolation of compounds.** Dried, milled root (3 kg) was extracted in 100 g portions with C<sub>6</sub>H<sub>6</sub> for 24 hr. Combined C<sub>6</sub>H<sub>6</sub> soln was concentrated to give a dark brown syrup (43 g) which was chromatographed on Si gel (170–230 mesh) with C<sub>6</sub>H<sub>6</sub>–MeOH (99:1, v/v). Eight crude fractions were collected and rechromatographed as follows. Fraction 2: Column chromatography in C<sub>6</sub>H<sub>6</sub>–Me<sub>2</sub>CO (95:5) and preparative TLC in hexane–C<sub>6</sub>H<sub>6</sub>–EtOAc (5:4:0.5), *R<sub>f</sub>* 0.29, gave neobanol (4) as colourless needles, mp 244–246°;  $\tau$  (Py-d<sub>3</sub>) 2.17, s, 1-H; 2.27, d, J2.2 Hz, 2'-H; 3.24, d, J2.2 Hz, 3'-H; 2.79, br. s, 4-H + 7-H; 3.40, s, 10-H; 4.07, s, 11a-H; 4.10, s, OCH<sub>2</sub>O; 5.00, br. s, 6a-OH; 5.46, s,  $\delta_{\text{eq,ax}}$ -H. Fraction 7: Rotenonone (2) was crystallized from C<sub>2</sub>H<sub>4</sub>Cl<sub>2</sub> as bright yellow needles, mp 298–305° (decomp.),  $\tau$  (100 MHz) 1.02, s, 1-H; 1.85, br. d, J8, 11-H; 3.01, br. d, J8, 10-H; 3.12, s, 4-H; 4.55, br. q, J<sub>1</sub> 9, J<sub>2</sub> 17, 5'-H; 4.83, br. s, =CH<sub>2</sub>; 5.00, s, 4'-H; 5.98, 6.04, s,

2  $\times$  OMe; 8.17, s, Me; UV (MeOH)  $\lambda_{\text{max}}$  217, 265, 295 nm (log  $\epsilon$  4.48, 4.29, 4.19) after chromatography in C<sub>6</sub>H<sub>6</sub>–hexane–EtOAc (5:5:2) and TLC in the same solvent system, *R<sub>f</sub>* 0.49. Neobanol (1). TLC *R<sub>f</sub>* 0.15 in C<sub>6</sub>H<sub>6</sub>–hexane–EtOAc (6:4:0.5), was obtained as a light yellow oil, *M*<sup>+</sup> 382,  $\tau$  (CDCl<sub>3</sub>) 1.76, s, 11-H; 2.44, d, J2.2 Hz, 2'-H; 3.00, s, 8-H; 3.33, s, 1-H; 3.34, d, J2.2 Hz, 3'-H; 3.46, s, 4-H; 5.03, m, 6a-H; 5.37, m, 6'-H; 6.17, s, OMe; 6.24, s, OMe; 6.44, s, 12a-OMe; CD (MeOH)  $[\theta]_{200} + 34500$ ,  $[\theta]_{205} 0$ ,  $[\theta]_{235-34500}$ ,  $[\theta]_{250} 0$ ,  $[\theta]_{265} + 6320$ ,  $[\theta]_{300} + 8050$ ,  $[\theta]_{370} 0$ ,  $[\theta]_{380-2300}$ . Acetylation of fraction 8 gave after chromatography in CHCl<sub>3</sub>–Et<sub>2</sub>O (97:3) and preparative TLC *R<sub>f</sub>* 0.56 in hexane–C<sub>6</sub>H<sub>6</sub>–EtOAc (5:5:0.5), 12a-acetoxyisomilletonone (3b) as light brown oil, *M*<sup>+</sup> 436,  $\tau$  (CDCl<sub>3</sub>) 2.09, d, J8, 11-H; 3.08, s, 1-H; 3.44, d, J8 10-H; 3.50, s, 4-H; 4.07, br. s, OCH<sub>2</sub>O; 4.51, m, 6a-H; 4.65, m, 5'-H; 5.53, m, 6-H; 6.92, m, 4'-H; 7.85, s, 12a-OAc; 8.23, br. s, 8'-Me; UV, (EtOH)  $\lambda_{\text{max}}$  210, 240, 300 nm (4.42, 4.08, 4.01), CD (MeOH)  $[\theta]_{220-38368}$ ,  $[\theta]_{240} 0$ ,  $[\theta]_{245} + 49704$ ,  $[\theta]_{252} 0$ ,  $[\theta]_{280-43600}$ ,  $[\theta]_{293} 0$ ,  $[\theta]_{300} + 26160$ ,  $[\theta]_{310} 0$ ,  $[\theta]_{325-68016}$ ,  $[\theta]_{390} 0$ .

**Rotenonone (2).** Dehydrorotenone (50 mg) in glacial HOAc (10 ml) was cooled to –5°. The reaction mixture was stirred for 1 hr at 0° after *n*-amyl nitrite (1.5 ml) was added after which it was allowed to warm to room temp. The mixture was extracted with C<sub>6</sub>H<sub>6</sub>, dried (Na<sub>2</sub>SO<sub>4</sub>) and evaporated to dryness to give after crystallization from C<sub>2</sub>H<sub>4</sub>Cl<sub>2</sub> (2) identical with the natural product.

**12-Acetoxyisomilletonone (3b).** Acetylation was done with Ac<sub>2</sub>O and dry pyridine according to standard procedure.

**Dehydroisomilletonone (7).** (3b) (20 mg) in EtOH–10% H<sub>2</sub>SO<sub>4</sub> (20 ml) was heated under reflux for 2 hr. Extraction with Et<sub>2</sub>O gave, after preparative TLC *R<sub>f</sub>* 0.29 in hexane–C<sub>6</sub>H<sub>6</sub>–EtOAc (6:5:1), dehydroisomilletonone (7) as a yellow oil,  $\nu_{\text{max}}$  CHCl<sub>3</sub> 1675 cm<sup>–1</sup> (unsat. C=O);  $[\alpha]_D^{236}$ °; UV EtOH  $\lambda_{\text{max}}$  210, 237, 295 nm (4.50, 4.13, 4.18);  $\tau$  (CDCl<sub>3</sub>) 2.12, d, J8.5, 11-H; 3.40, s, 1-H; 3.42, d, J 8.5, 10-H; 3.50 s, 4-H; 4.12, s, OCH<sub>2</sub>O; 4.80, m, 5'-H; 5.02, m, 7'-CH<sub>2</sub>; 8.22, br. s, 8'-Me; 5.43, br. s, 4'-CH<sub>2</sub> + 6-CH<sub>2</sub>; 6.83, m, 4'-H.

**Neoduleen (8).** Neobanol (20 mg) was dehydrated in a similar way as (3b) to give (8) as white needles (14 mg) from MeOH, mp 222–223°; *M*<sup>+</sup> 306; UV  $\lambda_{\text{max}}$  224, 252, 294, 346, 364 nm (4.30, 4.16, 3.84, 4.28, 4.18);  $\tau$  (CDCl<sub>3</sub>) 2.33, s, 1-H; 2.90, s, 7-H; 3.23, s, 10-H; 2.94, s, 4-H; 2.47, d, J2.2, 2'-H; 3.30, s, J2.2 Hz, 3'-H; 3.98, s, OCH<sub>2</sub>O; 4.45, s, 6-CH<sub>2</sub>.

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